

HO-1/BVR-A System Analysis in Plasma from Probable Alzheimer's Disease and Mild Cognitive Impairment Subjects: A Potential Biochemical Marker for the Prediction of the Disease

Fabio Di Domenico^a, Eugenio Barone¹, Cesare Mancuso^b, Marzia Perluigi^c, Annalisa Cocciolo^b, Patrizia Mecocci^c, D. Allan Butterfield^d and Raffaella Cocciola^a

^aDepartment of Biochemical Sciences, Sapienza University of Rome, Rome, Italy

^bInstitute of Pharmacology, Catholic University School of Medicine, Rome, Italy

^cInstitute of Gerontology and Geriatrics, University of Perugia, Perugia, Italy

^dDepartment of Chemistry, Center of Membrane Sciences, and Sanders-Brown Center on Aging, University of Kentucky, Lexington, KY, USA

Handling Associate Editor: Kenneth Hensley

Accepted 30 May 2012

Abstract. Several studies showed increased oxidative and nitrosative stress in plasma from patients with Alzheimer's disease (AD), however, little and controversial knowledge has emerged about the antioxidant functionality of the heme oxygenase-1/biliverdin reductase-A (HO-1/BVR-A) system in blood. The current study reports increased levels of both HO-1 and BVR-A in plasma from probable AD patients, as a result of the increased oxidative environment. However, the increase of oxidative stress in plasma result also in the increase of BVR-A 3-nitrotyrosine levels and the decrease of BVR-A phosphotyrosine levels and reductase activity, suggesting that nitrosative stress play the prominent oxidative role in plasma during AD. Our data on HO-1/BVR-A status in plasma closely correlate with recent reports in hippocampus of subjects with AD and arguably its early form, mild cognitive impairment. Moreover, we show that alterations on HO-1/BVR-A system are tightly connected with cognitive decline indexed by Mini-Mental Status Exam scores. We hypothesize that the HO-1/BVR-A system status in plasma might reflect the ongoing situation in the brain, offering an important biochemical tool for the potential prediction of AD at the earliest stages of the disease.

Keywords: Alzheimer disease's, bilirubin, biliverdin reductase-A, heme oxygenase-1, oxidative/nitrosative stress

Supplementary data available online: <http://www.j-alz.com/issues/32/vol32-2.html#supplementarydata06>

¹Current address: Brain Mind Institute, School of Life Sciences
Swiss Federal Institute of Technology (EPFL) Station 15, Lausanne,
Switzerland.

Correspondence to: Fabio Di Domenico, Department of Biochemical Sciences "A. Rossi Fanelli", Sapienza University of Rome, P.le Aldo Moro, Rome 5 00185, Italy. Tel.: +39 0649910900; Fax: +39 064440062; E-mail: fabio.didomenico@uniroma1.it.

INTRODUCTION

Alzheimer's disease (AD) is one of the most disabling dementing disorders in the elderly affecting about 20 million people worldwide, a number that is expected to increase since the average human life span

the ongoing situation in the brain. A thorough analysis of the HO-1/BVR-A quantity and post-translational modifications (PTMs) in blood-related bio fluids might offer an important biochemical tool for the prediction of AD, even in the early stages of the disease, and might represent a potential window of pharmacological intervention.

MATERIALS AND METHODS

Subjects

For this study, 36 subjects (12 with probable AD (pAD), 12 with MCI, and 12 cognitively normal persons) were enrolled. Complete clinical and neuropsychological evaluation was performed at the Department of Gerontology and Geriatrics, University of Perugia. Diagnosis of pAD was according to the National Institute of Neurological and Communicative Disease and Stroke and Alzheimer's disease (NINCDS-ADRDA) criteria [35]. MCI diagnosis was made following components of Petersen criteria [36]. Cognitively normal subjects (CTR) were enrolled among relatives of patients and subjects admitted to the Day Hospital of the same department for routine evaluation of the health status. All subjects underwent a thorough clinical, neurological, and neuropsychological evaluation; the neuropsychological assessment included the Mini Mental State Examination (MMSE), while the Clinical Dementia Rating scale measured the severity of dementia. The functional ability of subjects enrolled was evaluated on the basis of the activities of daily living and the instrumental activities of daily living (IADL). Subjects anxiety or depression (evaluated by the Geriatric Depression Scale), Hachinski ischemic score, cholesterol, erythrocytes sedimentation, and C-reactive protein was taken in consideration. The investigation conforms to the principles outlined in

the Declaration of Helsinki. All the subjects gave the written informed consent for blood donation. Blood was immediately centrifuged and plasma stored at -80°C until analyses. Subjects' demographic and clinical data are reported in Table 1 and Supplementary Table 1.

Samples preparation

Plasma (100 l) samples from each subject were depleted of the major plasma proteins (albumin and Immunoglobulins G) using PROT-BA depletion kit according to manufacturer instructions (Sigma-Aldrich, St. Louis, MO, USA). Protein concentration after albumin and Immunoglobulins G depletion was determined by using the Comassie protein assay (Pierce, Rock90(ceAr(j(Rock9ins)-32ock9ins)-331tx5dW6.2(alb)20k

nitrogen. The precipitate was re-suspended in 100 μ l DMSO used for the investigation. A Waters 600 and a Waters 996 Photodiode (Waters Corporation, Milford, MA, USA) were used for HPLC analysis. The column used was a Hyper BDS column C18, 250 mm length and 4.6 mm diameter \times 5 μ m. Solution A was prepared with acetonitrile/water 80/20, while solution B was prepared with ammonium acetate 0.1 mM/acetonitrile 80/20. The column was equilibrated with A/B 95/5 for 5 min, followed by linear gradient of B increase from 5 to 80 in 20 min, and a final equilibration step with A/B 95/5. The flow rate was 1 ml/min. Bilirubin detection was conducted at 450 nm. A calibration curve, using bilirubin IX alpha standard, was built before the plasma analysis.

Statistical analysis

Data are expressed as mean \pm SD of n independent samples. All statistical analyses were performed using a non-parametric one-way ANOVA with post hoc t-test. $p < 0.05$ was considered significantly different from control. Pearson correlations were calculated to test the linear association between cognitive test scores and markers of oxidative damage.

RESULTS

HO-1 protein quantitation and PTMs in plasma from pAD and MCI patients

The first parameter taken in consideration in the analysis of HO-1/BVR-A system in plasma of MCI and

pAD patients was the measurement of HO-1 protein quantity. Previous studies from our laboratory showed an increase of HO-1 in hippocampus of MCI and pAD subjects in response to environmental increased oxidative/nitrosative stress [23–25]. Although several authors described increased oxidative stress at the systemic level (both in plasma/serum and CSF) in AD subjects [37–41], data about the systemic HO-1 expression are still controversial [20, 27–29].

In the current study, plasma samples from three groups of patients, namely pAD, MCI, and age-matched CTR (12 samples each) were assessed for the analysis of HO-1/BVR-A quantity. Each sample was previously depleted for the most abundant proteins, IgG and albumin. As shown in Fig. 1A, HO-1 quantity significantly increased in both MCI and pAD compared to CTR samples. Interestingly, the trend of increase paralleled the severity of the pathology with MCI having 70% and AD 130% increase compared to CTR (Fig. 1A). After the expression evaluation, each sample was tested for protein specific PTMs. Previous studies from the Butterfield laboratory revealed that increased quantity of HO-1-bound HNE and HO-1 protein carbonyls was present in hippocampi from AD and MCI subjects, respectively [26]. Furthermore, in the same samples, increased quantity of pSer/Thr-HO-1 was detected in AD, suggesting impaired functionality of the protein in pathological conditions. In the current study we found no significant alterations in either HO-1 oxidation (Supplementary Figure 1) or HO-1 Ser/Thr phosphorylation in pAD/MCI plasma. Our results indicated that, in contrast with the hippocampus, plasma HO-1 might preserve its activity under severe pathologic conditions.

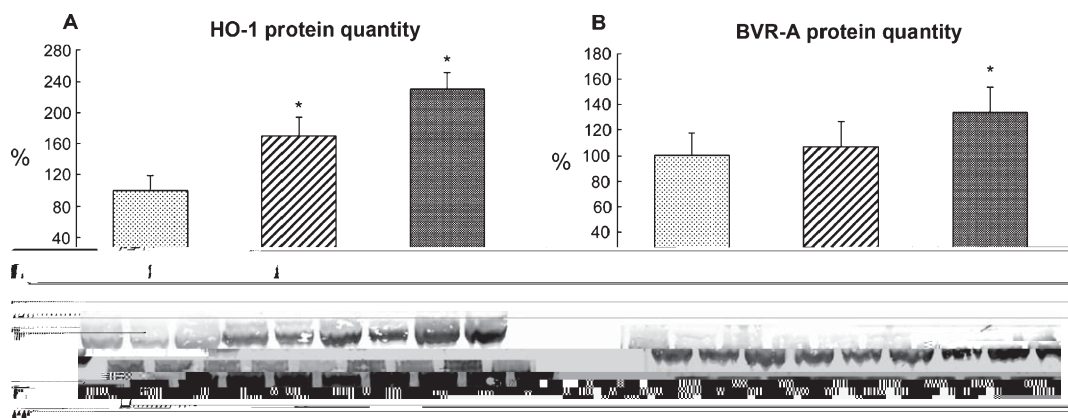


Fig. 1. HO-1 (A) and BVR-A (B) quantity in CTR, MCI, and pAD plasma samples. Densitometric values shown in the histograms are given as percentage of CTR, set as 100%. Data are expressed as mean \pm SD of 12 individual samples per group. $p < 0.05$.

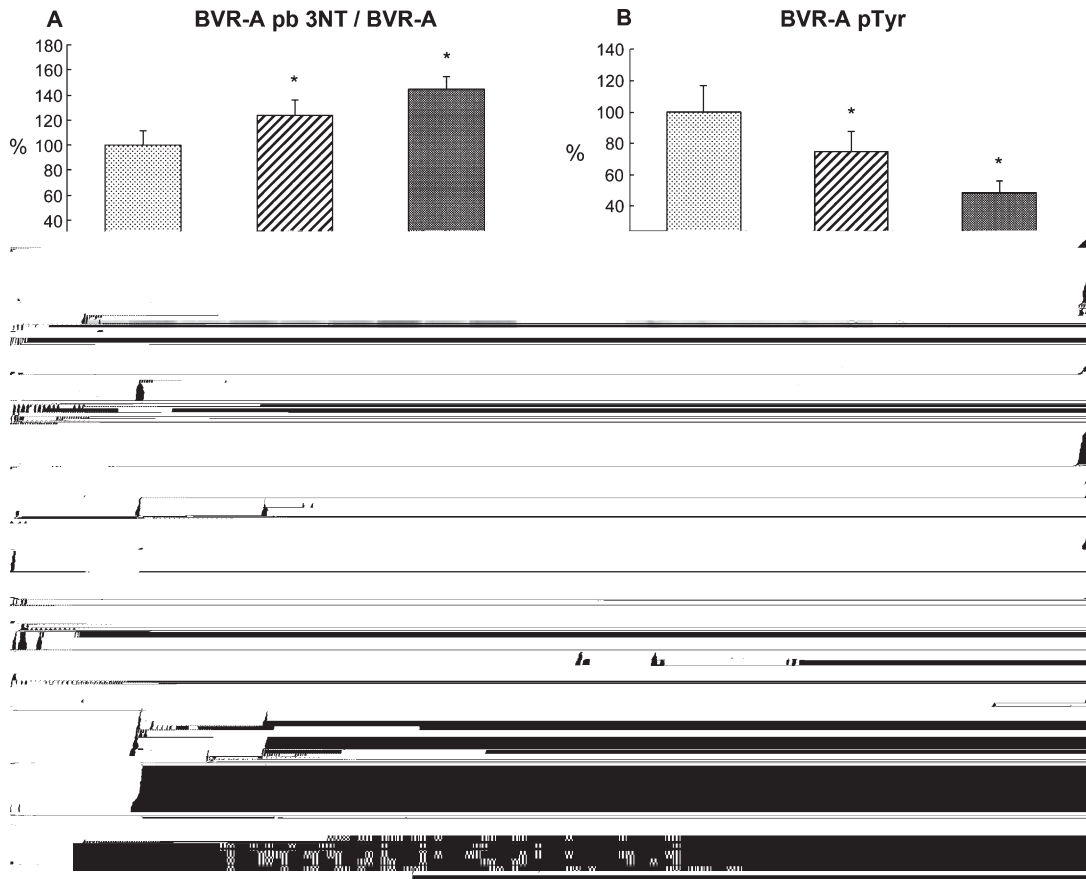


Fig. 2. BVR-A protein bound-3NT (A), phosphotyrosine (B), phosphoserine/threonine (C), and reductase activity (D) quantity in CTR, MCI, and pAD plasma samples. All the data from PTMs quantitation are normalized for BVR-A protein quantity. Densitometric values presented in the histograms are given as percentage of Control, set as 100%. Data are expressed as mean \pm SD of 2 individual samples per group. $p < 0.05$.

BVR-A protein quantitation, PTMs, and activity in plasma from pAD and MCI patients

Similarly to HO-1, BVR-A protein quantity increased in both pAD and MCI samples, but the induction of expression was slightly lower. Indeed, we observed an increase of 10% in MCI (not significant) and of 36% in pAD compared to CTR (Fig. 1B). Previous studies from the Butterfield laboratory showed increased BVR-A levels in the hippocampus of AD and MCI subjects, however, it was also shown that BVR-A underwent nitrosative stress modifications during AD and MCI exhibiting altered quantity of protein carbonyls and protein nitration [24]. As a consequence of the increased oxidative and nitrosative environment in AD that resulted in increased BVR-A oxidation and nitration, we also found a decrease



Fig. 4. Correlation graphs. A positive correlation ($p < 0.01, r = 0.53$) is shown for HO-1 and BVR-A protein quantity of pAD patients compared to CTR plasma (A) and for BVR-A protein quantity with BVR-A nitration in pAD compared to CTR ($p < 0.01, r = 0.63$) (B). Negative correlations were found for BVR-A protein quantity with BVR-A pTyr ($p < 0.01, r = -0.53$) (C) and BVR-A nitration with BVR-A pTyr ($p < 0.01, r = -0.73$) (D) in pAD versus CTR samples. A negative correlation ($p < 0.05, r = -0.48$) was found for BVR-A nitration with BVR-A pTyr in MCI compared to CTR plasma samples (E).

is corroborated by the parallelism between the changes occurring in plasma and those in the brain [23, 24, 26] and the correlation between plasma alteration and cognitive decline.

The main role of oxidative and nitrosative stress in the pathogenesis of AD has been largely investi-

An intriguing aspect of our previous research was the finding that the HO-1/BVR-A axis undergoes important PTMs in AD and MCI hippocampi [23–26]. This evidence obtained by studying an important brain area, such as the hippocampus, whose involvement in AD occurs at the earlier stages, prompted us to investigate whether or not the same PTMs were detected in

from the brain, passing through the blood brain barrier and released into the peripheral system (Scheme 1). With regard to BVR-A protein quantity, we previously stated that the investigation of only BVR-A expression might be ambiguous with regards to BVR-A functionality. Indeed, the analysis of BVR-A PTMs and reductase activity provide deeper and more detailed information about its real status during AD pathology [23, 24]. Consistent with the hippocampus results, our current findings demonstrate increased amount of protein bound 3-NT in pAD and MCI plasma, while no alterations of protein carbonylation and protein bound-HNE were detected. These results confirm the prevalence of nitrosative stress-induced modifications than protein carbonylation or protein bound-HNE on BVR-A structure during AD, as already observed in hippocampus [23].

a8ed8(408us)-1408uCTR3240749mo8(408us)-3r08uHO-3results,

- [7] Pratico D (2008) Oxidative stress hypothesis in Alzheimer's disease: A reappraisal *Trends Pharmacol Sci* 29, 609-615.
- [8] Markesbery WR (1997) Oxidative stress hypothesis in Alzheimer's disease *Free Radic Biol Med* 23, 134-147.
- [9] Sultana R, Butterfield DA (2010) Role of oxidative stress in the progression of Alzheimer's disease *Alzheimers Dis* 9, 341-353.
- [10] Butterfield DA, Perluigi M, Sultana R (2006) Oxidative stress in Alzheimer's disease brain: New insights from redox proteomics *Eur J Pharmacol* 545, 39-50.
- [11] Perluigi M, Sultana R, Cenini G, Di Domenico F, Memo M, Pierce WM, Coccia R, Butterfield DA (2009) Redox proteomics identification of 4-hydroxynonenal-modified brain proteins in Alzheimer's disease: Role of lipid peroxidation

- Subcommittee of the American Academy of Neurology 56, 1133-1142.
- [37] Zafrilla P, Mulero J, Xandri JM, Santo E, Caravaca G, Morillas JM (2006) Oxidative stress in Alzheimer patients in different stages of the disease. *Curr Med Chem* 3, 1075-1083.
- [38] Grossman M, Farmer J, Leight S, Work M, Moore P, Van Deerlin V, Pratico D, Clark CM, Coslett HB, Chatterjee A, Gee J, Trojanowski JQ, Lee VM (2005) Cerebrospinal fluid profile in frontotemporal dementia and Alzheimer's disease. *Ann Neurol* 57, 721-729.
- [39] Dildar K, Sinem F, Gokhan E, Orhan Y, Filiz M (2010) Serum nitrosative stress levels are increased in Alzheimer disease but not in vascular dementia. *Alzheimer Dis Assoc Dis* 24, 194-197.
- [40] Korolainen MA, Nyman TA, Nyyssonen P, Hartikainen ES, Pirttila T (2007) Multiplexed proteomic analysis of oxidation and concentrations of cerebrospinal fluid proteins in Alzheimer disease. *Clin Chem* 53, 657-665.
- [41] Skoumalova A, Ivica J, Santorova P, Topinkova E, Wilhelm J (2011) The lipid peroxidation products as possible markers of Alzheimer's disease in blood. *Exp Gerontol* 46, 38-42.
- [42] Aluise CD, Robinson RA, Beckett TL, Murphy MP, Cai J, Pierce WM, Markesbery WR, Butterfield DA (2010) Preclinical Alzheimer disease: Brain oxidative stress, Abeta peptide and proteomics. *Neurobiol Dis* 39, 221-228.
- [43] Butterfield DA, Stadman ER (1997) Protein oxidation processes in aging brain. In *Advances in Cell Aging and Gerontology* Timiras PS, Bittar EE, Eds. Elsevier B.V., pp. 161-191.
- [44] Montine TJ, Quinn JF, Milatovic D, Silbert LC, Dang T, Sanchez S, Terry E, Roberts LJ 2nd, Kaye JA, Morrow JD (2002) Peripheral F2-isoprostanes and F4-neuroprostanes are not increased in Alzheimer's disease. *Ann Neurol* 52, 175-179.
- [45] Irizarry MC, Yao Y, Hyman BT, Growdon JH, Pratico D (2007) Plasma F2A isoprostane levels in Alzheimer's and Parkinson's disease. *Neurodegener Dis* 4, 403-405.
- [46] Mufson EJ, Leurgans S (2010) Inability of plasma and urine F2A-isoprostane levels to differentiate mild cognitive impairment from Alzheimer's disease. *Neurodegener Dis* 7, 139-142.
- [47] Padurariu M, Ciobica A, Hritcu L, Stoica B, Bild W, Stefanescu C (2010) Changes of some oxidative stress markers in the serum of patients with mild cognitive impairment and Alzheimer's disease. *Neurosci Lett* 469, 6-10.
- [48] Pratico D, Clark CM, Lee VM, Trojanowski JQ, Rokach J, Fitzgerald GA (2000) Increased 8,12-iso-iPF2alpha-VI in Alzheimer's disease: Correlation of a noninvasive index of lipid peroxidation with disease severity. *Ann Neurol* 48, 809-812.
- [49] Pratico D, MY Lee V, Trojanowski JQ, Rokach J, Fitzgerald GA (1998) Increased F2-isoprostanes in Alzheimer's disease: Evidence for enhanced lipid peroxidation *in vivo*. *FASEB J* 12, 1777-1783.
- [50] Choi J, Malakowsky CA, Talent JM, Conrad CC, Gracy RW (2002) Identification of oxidized plasma proteins in Alzheimer's disease. *Biochem Biophys Res Commun* 293, 1566-1570.
- [51] Conrad CC, Marshall PL, Talent JM, Malakowsky CA, Choi J, Gracy RW (2000) Oxidized proteins in Alzheimer's plasma. *Biochem Biophys Res Commun* 275, 678-681.
- [52] Yu HL, Chertkow HM, Bergman H, Schipper HM (2003) Aberrant profiles of native and oxidized glycoproteins in Alzheimer plasma. *Proteomics* 3, 2240-2248.
- [53] Korolainen MA, Pirttila T (2009) Cerebrospinal fluid, serum and plasma protein oxidation in Alzheimer's disease. *Acta Neurol Scand* 119, 32-38.
- [54] Kim TS, Pae CU, Yoon SJ, Jang WY, Lee NJ, Kim JJ, Lee SJ, Lee C, Paik IH, Lee CU (2006) Decreased plasma antioxidants in patients with Alzheimer's disease. *Acta J Geriatr Psychiatry* 21, 344-348.
- [55] Mancuso C, Bonsignore A, Capone C, Di Stasio E, Pani G (2006) Albumin-bound bilirubin interacts with nitric oxide by a redox mechanism. *Antioxid Redox Signal* 8, 487-494.
- [56] Stocker R, Yamamoto Y, McDonagh AF, Glazer AN, Ames BN (1987) Bilirubin is an antioxidant of possible physiological importance. *Science* 235, 1043-1046.
- [57] Stocker R, Glazer AN, Ames BN (1987) Antioxidant activity of albumin-bound bilirubin. *Proc Natl Acad Sci U S A* 84, 5918-5922.
- [58] Dore S, Takahashi M, Ferris CD, Zakhary R, Hester LD, Guastella D, Snyder SH (1999) Bilirubin, formed by activation of heme oxygenase-2, protects neurons against oxidative stress injury. *Proc Natl Acad Sci U S A* 96, 2445-2450.
- [59] Barone E, Trombino S, Cassano R, Sgambato A, De Paola B, Di Stasio E, Picci N, Preziosi P, Mancuso C (2009) Characterization of the S-denitrosylating activity of bilirubin. *Cell Mol Med* 13, 2365-2375.
- [60] Kapitulnik J (2004) Bilirubin: An endogenous product of heme degradation with both cytotoxic and cytoprotective properties. *Mol Pharmacol* 66, 773-779.
- [61] Manolio TA, Burke GL, Savage PJ, Jacobs DR Jr, Sidney S, Wagenknecht LE, Allman RM, Tracy RP (1992) Sex- and race-related differences in liver-associated serum chemistry tests in young adults in the CARDIA study. *Clin Chem* 38, 1853-1859.
- [62] White GL Jr, Nelson JA, Pedersen DM, Ash KO (1981) Fasting and gender (and altitude?) in uence reference intervals for serum bilirubin in healthy adults. *Clin Chem* 27, 1140-1142.
- [63] Hale WE, Stewart RB, Marks RG (1983) Haematological and biochemical laboratory values in an ambulatory elderly population: An analysis of the effects of age, sex and drug use. *Ageing* 12, 275-284.
- [64] Ishizuka K, Kimura T, Yoshitake J, Akaike T, Shono M, Takamatsu J, Katsuragi S, Kitamura T, Miyakawa T (2002) Possible assessment for antioxidant capacity in Alzheimer's disease by measuring lymphocyte heme oxygenase-1 expression with real-time RT-PCR. *Ann N Y Acad Sci* 977, 173-178.
- [65] Maes OC, Kravitz S, Mawal Y, Su H, Liberman A, Mehindate K, Berlin D, Sahlas DJ, Chertkow HM, Bergman H, Melmed C, Schipper HM (2006) Characterization of alpha1-antitrypsin as a heme oxygenase-1 suppressor in Alzheimer plasma. *Neurobiol Dis* 24, 89-100.